

Effect of Ovulation Induction on Ovarian Histologies in a Rat Model

Özlem ULUSOY¹, Davut GÜVEN¹, Yurdanur SÜLLÜ², Özgür ERDOĞAN¹, İsmail KILIÇOĞLU¹, Cazip ÜSTÜN¹, İdris KOÇAK¹

Samsun, Turkey

OBJECTIVE: To examine the effect of human menopausal gonadotropin(HMG) and follitropinBeta (rFSH) on the ovarian histologies in a rat model.**STUDY DESIGN:**Thirty-nine female, one-year old rats were enrolled for the trial. They were divided into three groups. In the first group, 13 rats received, six cycles of ovulation induction with human menapausal gonadotropin.In the second group, 13 rats received six cycles of ovulation induction with follitropin beta the third study group consisted of 13 rats which received six cycles of saline intramuscularly.**RESULT:** The mean number of the cells that stained positive for Ki-67 was 42.3±20.6, 44.9±27.3and 42.5±24.8 in the follicles, respectively.The mean number of cells that stained positive for Ki-67 in epithelium was 0.15±0.42, 0.04±0.14,0.05±0.18, respectively. The mean dysplasia score was 2.46±2.10, 1.69±1.13 and 1.62±1.89 in the ovarian epithelial cells respectively.**CONCLUSION:**Development of malignant lesion were not found in any of the rat ovaries after ovulation induction. As a result of this study, we found out that human menapausal gonadotropin and follitropin beta used in treatment of infertility, when administered for six cycles in accordance with the dosage determined, do not have the potential to develop neoplasia in rats.**Key Words:** Ovulation induction, Ki-67 expression, Dysplasia score*Gynecol Obstet Rebrod Med;15:1 (30 - 33)*

Introduction

In the 1980 s, case reports of ovarian cancer in women undergoing assisted conception raised concern about the long-term effects of infertility treatment¹ subsequent epidemiological studies reported positive associations between exposure to fertility drug used to stimulate ovulation and the risk of ovarian cancer.However, more recent studies have not confirmed this association, nor have they found an association and the overall risks of breast cancer.²

The Ki-67 antigen, which is coded by a gene on chromosome 10 is expressed in G1, S and G2 phases in cycling cell but not in the resting phase G0. The Ki-67 score partly correlates with other proliferation markers like percentage of S-phase cell and mitotic count.^{3,4}

High-grade ovarian carcinomas display higher Ki-67 expression.⁵

¹Department of Obstetrics and Gynecology and ²Pathology Scholl of Medicine Ondokuz Mayıs University, Samsun

Address of Correspondence: Davut Güven
Cumhuriyet Mah. Atatürk Bul. 11. Sok.
No:1 Bahadır Apt. D:6 Atakum
Samsun
dguven@omu.edu.tr

Submitted for Publication: 29.01.2009

Accepted for Publication: 20.02.2009

Our purpose in this study is to investigate the effect of ovulation induction on dysplasia score and Ki-67 expression of the ovarian epithelium in a rat model.

Material and Method

With the permission of the local ethic committee of the Ondokuz Mayıs University school of medicine, one-year-old and nearly 160-270 g weighing, 39 sexually mature Wistar-Albino female rats were enrolled in the study.Rats were divided into three groups. In the first group, rats received six cycles of ovulation induction with follitropin Beta.The third study group consisted of 13 rats which received six cycles of saline intramuscularly (im). Vaginal smears were obtained from both groups every morning for two consecutive cycles and 5-day cycles were divided into phases as estrous; diestrous days 1,2,3 and proestrous day. On estrous day 2 of the six cycle, group 1 received 150 IU/kg HMG im at 5 PM, group 2 received 150 IU/kg rFSH im at 5 PM, group 3 received saline intramuscularly. On the proestrous day of the third cycle, 75 IU/kg human chorionic gonadotropin (hCG). Was injected im at 5 PM in group 2 and Group 3 received saline injection on the proestrous day of the six cycle at 5 PM. Each ovulation induction cycle was repeated every 2 weeks in the three groups, given consideration to the fact that their menstrual cycle is 5 day long.

Laparotomy and bilateral oophorectomy were performed in all rats under ether anesthesia.Laparotomy was performed

at the seventh cycle, 2 days after the last human chorionic gonadotropin injection.

Laparotomy with midline incision to all the rats were done under the ketamine hydrochloride anesthesia (2 ml/kg). The rats were killed after their ovaries were extracted. Ovarian tissue was fixed with 10% formaldehyde for the histopathological examination. After the paraffin blocks were prepared, 5 µm thick cross sections were taken and six cross sections prepared from each ovary were dyed in hematoxylin and eosin. The preparations were examined by a pathologist.

In the histopathological examination, the following parameters have been investigated: a malignant lesion, ovarian cyst and its size, stratification of ovary epithelial cells, a local epithelial accumulation (tufting), a mitotic index in granulosa cells, polymorphism in epithelial cells and the chromatin intensity, nuclear atypia in ovarian cyst epithelium, mitotic activity, and papillary formation in the ovarian cyst epithelium. The most dysplastic region in the histopathological examination. The size of nucleus was measured with ocular micrometer.

Each induced group was compared with its control group, the Mann-Whitney U test and chi-square test, Kruskal-Wallis test and Mann-Whitney U-test were used.

Results

In group 1,2,3, the mean number of the cell that stained for Ki-67 was 42.3±20.6, 44.9±27.3, 42.5±24.8 in the follicles. Respectively (p>0.05, Table I).

	Comparison of Ki-67 immune reactivity of the follicular cell in three groups	P Value	Comparison of Ki-67 immune reactivity of the epithelium	P Value
Group 1 HMG	42.3±20.6	>0.65	0.15±0.42	>0.05
Group 2 rFSH	44.9±27.3	>0.05	0.04±0.14	>0.05
Group 3	42.5±24.8	>0.05	0.05±0.18	>0.05

The data are presented as mean±standard deviation

*Statistically significant (p<0.05), Mann-Whitney U test

The mean number of the cell that stained positive for Ki-67 in the epithelium, were not significantly in the ovulation induction group (0.15±0.42, 0.04±0.14, 0.05±0.18 respectively) compared to the control group p>0.005. The mean dysplasia score in the ovulation induction groups was not significantly (2.46±2.10, 1.69±2.13, 1.62±1.89 P>0.05 respectively)

The distribution of the histologic features of the dysplasia score is presented in table II.

Table II: Distribution of the histologic abnormalities in the three groups.

	Scoring of groups								
	Groups 1 n:13			Groups 2 n:13			Groups 3 n:13		
Histologic Features	1	2	3	1	2	3	1	2	3
Epithelial multilayerin	6	7	0	8	5	0	9	4	0
Tufting	12	1	0	13	0	0	13	0	0
Nuclear chromatin irregularity	12	1	0	13	0	0	13	0	0
Nuclear contour irregularity	12	1	0	13	0	0	13	0	0
Nuclear size	6	7	0	8	5	0	8	5	0
Nuclear/cytoplasmic ratio	6	7	0	8	5	0	8	5	0
Presence of nucleoli	13	0	0	11	2	0	13	2	0
Presence and number of mitotic figures	13	0	0	13	0	0	13	0	0

0: Normal, 1: Moderate abnormality, 2: Severe abnormality

There was not a positive correlation between the Ki-67 index and the dysplasia (P>0.05). Cell expressing Ki-67 identified in the epithelium, follicle (Fig I.II.III).

The follicular cells in many of the developing follicles of the ovulation induced ovaries not showed intense nuclear immunoreactivity compared to the controls (Table II).

When number of the Ki-67 stained cells in three groups are evaluated, results for group 1,2,3 are 1134.54±236.54, 1154.31±246.05 and 922.15±170.31 respectively. Not statistical significance was found between those three groups in terms of the follicles and surface epithelium Ki 67 index, ovarian dysplastic alteration and the average number of the Ki 67 stained cell. Not statistical significance was found between the individual evaluations of the groups

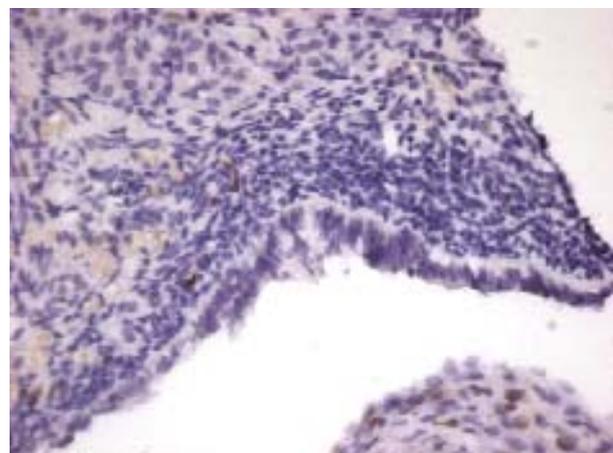


Figure 1: Ki-67 Immunostaining in the ovarian epithelium of group 1X100 (A), X 200 (B)

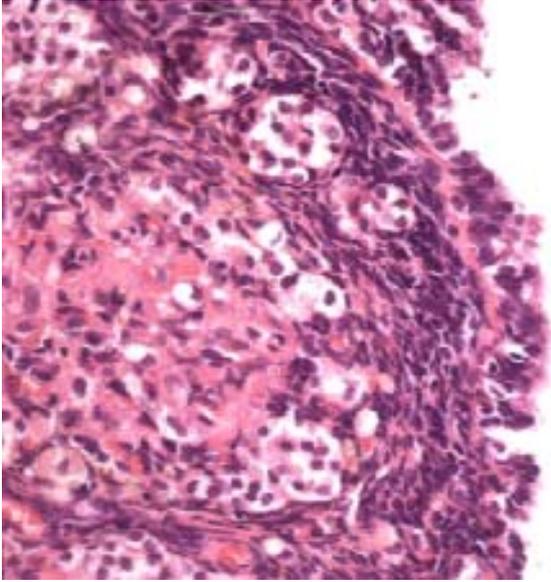


Figure II: Epithelial dysplasia in the rat ovarian epithelium in group 2. H-E, X 400

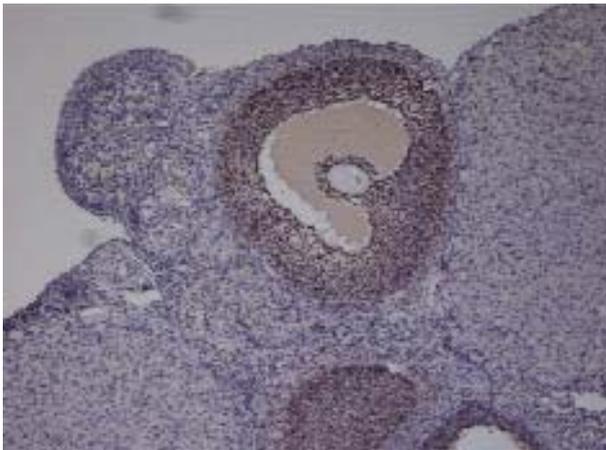


Figure III: Ki67 Immunostaining in the ovarian follicle of group 1X100 (A), X 200(B)

Discussion

Currently available data in the literature suggest that an association between ovulation induction and ovarian cancer does not indicate necessarily a causal effect infertility alone is an independent risk factor for the development of ovarian cancer. Nulliparous women with refractory infertility may harbor a particularly high risk of ovarian cancer, irrespective of their use of fertility drugs. Furthermore, the apparent association between fertility drug use and ovarian cancer may arise because these women are the most likely to have used ovulation stimulating agents as part of their infertility treatment.⁶ In 1971, Fathalla proposed that the etiology of ovarian cancer was related to "with each ovulation the ovarian surface (epithelium) was thought to incur minor trauma. The cumulative effect of repetitive surface injury was hypothesized to con-

tribute to the development of ovarian neoplasm.^{6,7}

Preliminary results of an ongoing case-control study in Italy were published recently and did not suggest a role for ovulation stimulation in the etiology of ovarian cancer.^{6,8}

Gonadotropins were also blamed for inducing ovarian cancer at high and prolonged doses, as used in ovulation induction. Gonadotropins can induce Cyclooxygenase 2 and increase prostaglandin production, which in turn causes the loss of basement membrane of the ovarian surface epithelium and alters the biology of the epithelial cells in cell contact signaling and organization.^{3,7} Our study is an experimental study using HMG and rFSH in rats has no noticeable effect on the ovary because the comparison of the groups 2,3 and controls did not show any pre-malign changes.

Corakci et al. Found increased Ki-67 expression in the ovulation induction group with increasing dysplasia score. This finding should be evaluated with caution as the ovaries were examined immediately after six cycles of ovulation induction increases epithelial dysplasia scores and Ki-67 expression in rat ovaries.^{3,9}

Celik et al. No malignant ovarian lesion was found in the ovulation induction. Ovarian cyst development was most frequent in the rats that underwent six cycles of ovulation induction.

Significant difference was found between induction and control group for cellular and nuclear polymorphism, presence of nucleolus, and nuclear chromatin density.¹⁰

In this study, neither the ovulation induced rats nor the control group rats were found to have neoplastic lesion (Ki-67 expression and dysplastic features).

The etiology of ovarian cancer is multifactorial, and the genetic, environmental, and endocrinologic factors or indirectly. Therefore, large prospective studies consisting of meticulously selected proper control groups are needed.

Ovulasyon İndüksiyonunun Over Histolojisi Üzerine Etkilerinin Rat Modelinde İncelenmesi

AMAÇ: Bu çalışmanın amacı overian histoloji üzerine ovulasyon indüksiyonunu etkilerinin rat modelinde incelenmesidir. Bu amaçla otuz dokuz adet bir yaşında dişi rat kullanıldı.

GEREÇ ve YÖNTEM: Ratlardan üç grup oluşturuldu. İlk gruptaki 13 rata ovulasyon indüksiyonu için altı siklus human menopozal gonadotropin ikinci gruptaki 13 rata 6 siklus follitropin beta verildi. Üçüncü gruptaki 13 rata intramusküler olarak altı siklus salin uygulandı. Altı siklus sonra bilateral oofektomi uygulandı.

BULGULAR: Over epitelinin displazi scoru ve Ki-67 ekspresyonu incelendi. Ortalama olarak folliküllerde Ki-67 pozitif saptanan hücre sayıları sırasıyla 42.3 ± 20.6 , 44.9 ± 27.3 ve

42.5±24.8 olarak bulundu. Epitelde Ki-67 pozitif saptanan hücre sayıları ortalama olarak sırasıyla 0.15±0.42, 0.04±0.14, 0.05±0.18 olarak saptandı. Over epitelindeki ortalama displazi skoru sırasıyla 2.46±2.10, 1.69±1.3 and 1.62±1.89 olarak saptandı. Ovulasyon indüksiyonu sonrasında hiçbir rat overinde malign lezyon gelişimi saptanmadı.

SONUÇ: Rat modelinde altı siklus boyunca tanımlanan dozlarda kullanılan human menopozal gonodotropin ve folitropin beta neoplazi gelişimi için potansiyel sebep oluşturmamıştır.

Anahtar Kelimeler: Ovulasyon indüksiyonu, Ki-67, Displazi skoru

References

1. Fishel S, and Jackson P. follicular stilation for high tech pregnancies: are we playing it safe (?) Br. Med. J., 1980;299:309-11
2. Pat Doyle, Noreen Maconochie, Valeries Beral, Anthony J. Swetdlow and S.L.Tan. Cancer incidence folowing treatment for infertility at a clinic in the UK. Human Rep. 2002;17 (8) 2209-2213.
3. Van Deist. PJ, Brugal G, Baak JPA. Proliferation markers in tumours; interpretation and clinical value. JC in Pathol 1998;51;716-24.
4. Corakcı A, Filiz S, Çalışkan E, Dalcık C, Özeren S, Dalcık H. The effects of ovulation induction on ovarian epithelium dysplasia scores and Ki 67 expression: an experimental study on rats. Int J Gynecol Cancer 2005;15:866-71
5. Korkolopoulousı, Konstantinidou AE et al. Combined evaluation of p27 kip1 and Ki 67 expression provides independent information on overall survival of ovarian carsinoma patiens. Gynecol oncol 2002;85:404-14.
6. Robert E. Bristow, Beth Y. karlan. Ovulation induction, infertility, and ovarian cancer risk. Fertility and Sterility. 1996;66:499-507.
7. Fathalla MF. Incessant ovulation a factor in ovarian neoplasia? Lancet. 1971;2:163.
8. Franceschi S, la Vecachia C, Negri E et al. Fertility drugs and risk of epithelial ovarian cancer in Italy. Hum Repred 1994;9:1673-5.
9. Smith ER, Xu X. Etiology of epithelial ovarian cancer: a celular mechanism fort he role of gonadotropins. Gynecol Oncol; 2003;91:1-2
10. Celik C, Gezginç K, Aktan A, Acar A et al. Effects of ovulation induction on ovarian morphology: an animal study. Int J Gynecol Cancer 2004;14:600-606.